



Human Bronchial Smooth Muscle Cells (HBSMC)

Catalog Number: 3400

Cell Specification

Airway smooth muscle is fundamental in maintain airway tone. Their specialized ultrastructural features and regulatory mechanisms are the basis for normal airway function. In patients with severe asthma, the characteristic feature is an increase in bronchial smooth muscle mass. This increased smooth muscle mass is related either to the abnormal bronchial smooth muscle cell proliferation or the accumulation of contractile protein. Airway smooth muscle cell proliferation has been the focus of considerable attention, as it is a quantitatively important component of the airway wall remodeling response in asthma and has been suggested as a suitable target for the development of novel anti-asthma agents. Airway smooth muscle cells express CT-1 [1] and release GM-CSF, RANTES and IL-8 *in vitro* [2]. In order to develop anti-remodeling drugs, researchers have utilized cell culture techniques to elucidate the cellular and molecular mechanisms underlying bronchial smooth muscle cell proliferation and to identify the critical cell cycle events that regulate the cell growth.

HBSMC from ScienCell Research Laboratories are isolated from human bronchi and bronchioles. HBSMC are cryopreserved at secondary culture and delivered frozen. Each vial contains $>5 \times 10^5$ cells in 1 ml volume. HBSMC are characterized by immunofluorescent method with antibodies to α -smooth muscle actin and desmin. HBSMC are negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi. HBSMC are guaranteed to further expand for 15 population doublings at the condition provided by ScienCell Research Laboratories.

Recommended Medium

It is recommended to use smooth muscle cell medium (SMCM, Cat. No. 1101) for the culturing of HBSMC *in vitro*.

Product Use

HBSMC are for research use only. It is not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

Storage

Directly and immediately transfer cells from dry ice to liquid nitrogen upon receiving and keep the cells in liquid nitrogen until cell culture needed for experiments.

Shipping

Dry ice.

Reference

- [1] Zhou, D., Zheng, X., Wang, L., Stelmack, G., Halayko, A. J., Dorscheid, D. and Bai, T. R. (2003) Expression and effects of cardiotrophin-1 (CT-1) in human airway smooth muscle cells. *British Journal of Pharmacology* 140:1237-1244.
- [2] Oltmanns, U., Issa, R., Sukkar, M. B., John, M. and Chung, K. F. (2003) Role of c-jun N-terminal kinase in the induced release of GM-CSF, RANTES and IL-8 from human airway smooth muscle cells. *British Journal of Pharmacology* 139:1228-1234.

Instruction for culturing cells

Caution: Cryopreserved cells are very delicate. Thaw the vial in a 37°C waterbath and return them to culture as quickly as possible with minimal handling!

Set up culture after receiving the order:

1. Prepare a poly-L-lysine coated flask ($2 \mu\text{g}/\text{cm}^2$, T-75 flask is recommended). Add 10 ml of sterile water to a T-75 flask and then add 15 μl of poly-L-lysine stock solution (10 mg/ml, ScienCell cat. no. 0413). Leave the flask in incubator overnight (minimum one hour at 37°C incubator).
2. Prepare complete medium: decontaminate the external surfaces of medium and medium supplements with 70% ethanol and transfer them to sterile field. Aseptically open each supplement tube and add them to the basal medium with a pipette. Rinse each tube with medium to recover the entire volume.
3. Rinse the poly-L-lysine coated flask with sterile water twice and add 20 ml of complete medium to the flask. Leave the flask in the hood and go to thaw the cells.
4. Place the vial in a 37°C waterbath, hold and rotate the vial gently until the contents are completely thawed. Remove the vial from the waterbath immediately, wipe it dry, rinse the vial with 70% ethanol and transfer it to a sterile field. Remove the cap, being careful not to touch the interior threads with fingers. Using a 1 ml eppendorf pipette gently resuspend the contents of the vial.
5. Dispense the contents of the vial into the equilibrated, poly-L-lysine coated culture vessels. A seeding density of 5,000 cells/cm² is recommended.

Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to the cells than the effect of DMSO residue in the culture. It is also important that smooth muscle cells are plated in poly-L-lysine coated flasks that promote cell attachment.

6. Replace the cap or cover of flask, and gently rock the vessel to distribute the cells evenly. Loosen cap if necessary to permit gas exchange.
7. Return the culture vessels to the incubator.
8. For best result, do not disturb the culture for at least 16 hours after the culture has been initiated. Change the growth medium the next day to remove the residual DMSO and unattached cells, then every other day thereafter. A healthy culture will display spindle shaped, usually in a homogeneous bundle or sheet of cells rather than scattered single cells and the cell number will be double after two to three days in culture.

Maintenance of Culture:

1. Change the medium to fresh supplemented medium the next morning after establishing a culture from cryopreserved cells. For subsequent subcultures, change medium 48 hours after establishing the subculture.
2. Change the medium every other day thereafter, until the culture is approximately 50% confluent.
3. Once the culture reaches 50% confluence, change medium every day until the culture is approximately 90% confluent.

Subculture:

1. Subculture the cells when they are over 90% confluent.
2. Prepare poly-L-lysine coated cell culture flasks (2 $\mu\text{g}/\text{cm}^2$).
3. Warm medium, trypsin/EDTA solution, trypsin neutralization solution, and DPBS to **room temperature**. We do not recommend warming the reagents and medium at 37°C waterbath prior to use.
4. Rinse the cells with DPBS.
5. Incubate cells with 10 ml of trypsin/EDTA solution (in the case of T-75 flask) until 80% of cells are rounded up (monitored with microscope). Add 10 ml of trypsin neutralization solution to the digestion immediately and gently rock the culture vessel.

Note: Use ScienCell Research Laboratories' trypsin/EDTA solution that is optimized to minimize the killing of the cells by over trypsinization.

6. Harvest and transfer released cells into a 50 ml centrifuge tube. Rinse the flask with another 10 ml of growth medium to collect the residue cells. Examine the flask under microscope to make sure the harvesting is successful by looking at the number of cells left behind. There should be less than 5%.
7. Centrifuge the harvested cell suspension at 1000 rpm for 5 min and resuspend cells in growth medium.
8. Count cells and plate them in a new, poly-L-lysine coated flask with cell density as recommended.

Caution: Handling human derived products is potentially biohazardous. Although each cell strain testes negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate, therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].

[1]. Grizzle, W. E., and Polt, S. S. (1988) Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues. J Tissue Culture Methods. 11(4).