



Human Calvarial Osteoblasts (HCO)

Catalog Number: 4600

Cell Specification

Bone is a dynamic tissue, being continuously remodeled by the coordinated actions of osteoclasts and osteoblast lineage. Osteoblasts, the bone-forming cells, are derived originally from pluripotent mesenchymal stem cells. They synthesize and secrete organic extracellular matrix, osteoid, which is composed primarily of type I collagen. Osteoid is calcified by osteoblasts and during this process the cells become encased in lacunae within the calcified material and become osteocytes. Osteoblasts express protease-activated receptor-1 and vesicular endothelial cell growth factor [1]. Studies show that Leukemia inhibitory factor can bind to the osteoblast cell surface and induce bone formation both *in vitro* and *in vivo* [2]. The balance between osteoblast recruitment, proliferation, differentiation and apoptosis in sutures between cranial bones is essential for calvarial bone formation [3].

HCO from ScienCell Research Laboratories are isolated from human calvariae. HCO are cryopreserved at primary cultures and delivered frozen. Each vial contains $>5 \times 10^5$ cells in 1 ml volume. HCO are characterized by the cytochemically detection of AP and mineral deposition. HCO are negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi. HCO are guaranteed for 15 population doublings at the conditions provided by ScienCell Research Laboratories.

Recommended Medium

It is recommended to use Osteoblast Medium (OsM, Cat. No. 4601) for the culturing of HCO *in vitro*.

Product Use

HCO are for research use only. It is not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

Storage

Directly and immediately transfer cells from dry ice to liquid nitrogen upon receiving and keep the cells in liquid nitrogen until cell culture needed for experiments.

Shipping

Dry ice.

Reference

- [1] Steinbrech, D. S., Mehrara, B. J., Saadeh, P. B., Greenwald, J.A., Spector, J. A., Gittes, G. K. and Longaker, M. T. (2000) VEGF expression in an osteoblast-like cell line is regulated by a hypoxia response mechanism. *Am. J. Physiol. Cell Physiol.* 278: C853-C860.
- [2] Dazai, S., Akita, S., Hirano, A., Rashid, M. A., Naito, S., Akino, K., Fujii, T. (2000) Leukemia inhibitory factor enhances bone formation in calvarial bone defect. *J. Craniofac. Surg.* 11(6):513-20.
- [3] Marie, P. J., Debais, F., Hay, E. (2002) Regulation of human cranial osteoblast phenotype by FGF-2, FGFR-2 and BMP-2 signaling. *Histol. Histopathol.* 17(3):877-85.

Instruction for culturing cells

Caution: Cryopreserved cells are very delicate. Thaw the vial in a 37°C waterbath and return them to culture as quickly as possible with minimal handling!

Set up culture after receiving the ordering:

1. Place the vial in a 37°C waterbath, hold and rotate the vial gently until the contents are completely thawed. Remove the vial from the waterbath immediately, wipe it dry, and transfer it to a sterile field. Rinse the vial with 70% ethanol, and then wipe to remove excess. Remove the cap, being careful not to touch the interior threads with fingers. Using 1 ml eppendorf pipette gently resuspend the contents of the vial.
2. Dispense the contents of the vial into the equilibrated, poly-l-lysine-coated culture vessels (T-75 flask). A seeding density of $>5,000$ cells/cm² is recommended.

Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to cells than the effect of DMSO residue in the culture. It is also important that osteoblasts are plated in poly-l-lysine-coated culture vessels that promote osteoblast attachment.

3. Replace the cap or cover, and gently rock the vessel to distribute the cells evenly. Loosen caps if necessary to permit gas exchange.
4. Return the culture vessels to the incubator.
5. Change the growth medium 4 - 6 hours after plating to remove the residual DMSO and unattached cells, then every other day thereafter. A health culture will display normal cell morphology, nongranular cytoplasm, and the cell number will be doubled after two to three days in culture.

Subculture:

1. Subculture the cells when they are 90% confluent.
2. Prepare poly-L-lysine-coated cell culture vessels.
3. Warm medium, trypsin/EDTA solution, trypsin neutralization solution, and DPBS to room temperature.
4. Rinse the cells with DPBS.
5. Incubate cells with 5 ml of trypsin/EDTA solution (in the case of T-75 flask) until 80% of cells are rounded up (monitored with microscope).

6. Gently remove trypsin/EDTA solution from flask; hold the flask with one hand and use side of the flask to strike the open palm of the other hand. The motion, which should be parallel to the floor of the flask, will loosen the cells.

Note: Use ScienCell Research Laboratories' trypsin/EDTA solution that is optimized to minimize the killing of the cells by over trypsinization.

7. Add 10 ml of trypsin neutralization solution to the flask and gently rock the flask.
8. Harvest and transfer released cells into a 15 ml centrifuge tube. Rinse the flask with another 5 ml of trypsin neutralization solution to collect the residue cells. Examine the flask under microscope to make sure the harvesting is successful by looking at the number of cells left behind. There should be less than 5%.
9. Centrifuge the cell suspension at 1000 rpm for 5 min and resuspend cells in growth medium.
10. Count cells and plate them in a new, poly-L-lysine-coated flask with cell density as recommended.

Caution: Handling human derived products is potentially biohazardous. Although each cell strain testes negative for HIV, HBV and HCV DNA, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].

[1]. Grizzle, W. E., and Polt, S. S. (1988) Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues. J Tissue Culture Methods. 11(4).