



## Choline Assay (Cho)

*Cat. No. 8588  
100 Tests in 96-well plate*

### Introduction

Choline is an essential nutrient and plays a key role in many biological processes. Choline and its metabolites are needed for various physiological purposes, including: structural integrity of cell membranes, cell membrane signaling, cholinergic neurotransmission, and additionally, they are a major source of methyl groups that are needed for the biosynthesis of lipids and the regulation of metabolic pathways. Choline deficiency plays a role in liver disease, atherosclerosis, neurological disorders, and organ dysfunction. This colorimetric assay is based on choline oxidase-catalyzed oxidation of free choline, where the formed hydrogen peroxide is then catalyzed by peroxidase and reacts with 4-aminoantipyrine to form a colored product that can be read at 550nm. The intensity of the colored reaction product is directly proportional to the concentration of choline in the sample.

### Kit Components

Cat. No.	# of vials	Reagent	Quantity	Storage
8588a	1	Assay buffer	25 mL	4°C
8588b	1	Choline standard	1.0 mL	-20°C
8588c	1	Substrate mix	1.6 mL	-20°C
8588d	1	Enzyme mix	0.2 mL	-20°C

### Product Use

The Choline Assay kit measures choline level in various types of samples, including cells, serum, plasma, and tissue. This product is for research purposes only and is not approved for use in animals, humans, or diagnostic procedures.

### Quality Control

Data from the Choline Assay kit using choline solutions with concentrations ranging from 6.25-400  $\mu$ M show a linear relationship between OD<sub>550nm</sub> and choline concentration (Figure 1).

### Shipping

Shipped on dry ice.

## Procedures (96-well plate)

### A. Preparation of choline standard

1. Add 60  $\mu\text{L}$  of choline standard (8588b) to 15  $\mu\text{L}$  of assay buffer (8588a) to make a 75  $\mu\text{L}$  solution of 800  $\mu\text{M}$  choline.
2. Obtain 8 test tubes, add 50  $\mu\text{L}$  of assay buffer (8588a) into each tube and label them #1 through #8.
3. Add 50  $\mu\text{L}$  of the 800  $\mu\text{M}$  choline solution into tube #1 and mix well to obtain the 400  $\mu\text{M}$  choline standard.
4. Transfer 50  $\mu\text{L}$  of the 400  $\mu\text{M}$  choline standard from tube #1 to tube #2 and mix well to obtain the 200  $\mu\text{M}$  choline standard.
5. Repeat step 4 for tubes #3-7 to serially dilute the choline standards. Do not add any choline to tube #8, which serves as a blank.
6. Obtain a 96-well test plate, prepare 2 replicates (A and B) of each choline standard by aliquoting 20  $\mu\text{L}$ /well of each choline standard into duplicate wells of the 96-well test plate, following the plate format below:

	#1	#2	#3	#4	#5	#6	#7	#8
A	400 $\mu\text{M}$	200 $\mu\text{M}$	100 $\mu\text{M}$	50 $\mu\text{M}$	25 $\mu\text{M}$	12.5 $\mu\text{M}$	6.25 $\mu\text{M}$	Blank
B	400 $\mu\text{M}$	200 $\mu\text{M}$	100 $\mu\text{M}$	50 $\mu\text{M}$	25 $\mu\text{M}$	12.5 $\mu\text{M}$	6.25 $\mu\text{M}$	Blank

### B. Preparation of test samples

1. Cells or Tissues can be homogenized in 4 volumes of the assay buffer (8588a). Centrifuge the samples at 13,000  $\times g$  for 10 minutes to remove any insoluble material. The soluble fraction may be assayed directly.
2. Samples should be serially diluted to ensure that the readings are within the range of the standard curve. Prepare test samples to a final volume of 20  $\mu\text{L}$ /well in the 96-well flat bottom plate.

### C. Working reagent preparation and measurements

1. For each well to be assayed, prepare the working reagent by mixing 62  $\mu\text{L}$  assay buffer (8588a), 16  $\mu\text{L}$  substrate mix (8588c), and 2  $\mu\text{L}$  enzyme mix (8588d).
2. Add 80  $\mu\text{L}$  of the working reagent mix into each well of the 96-well plate containing choline standard, samples, and blank. Incubate the reaction for 60 minutes at room temperature in the dark.
3. Read the absorbance with an ELISA plate reader at 550 nm.

### D. Calculations

1. Subtract the  $\text{OD}_{550\text{nm}}$  value of the blank from the  $\text{OD}_{550\text{nm}}$  values of the standards and samples to obtain the  $\Delta\text{OD}_{550\text{nm}}$  value.
2. Based on the calibrated  $\Delta\text{OD}_{550\text{nm}}$  of the choline standard, make a standard curve by plotting  $\Delta\text{OD}_{550\text{nm}}$  as a function of choline concentration (see Figure 1 for a typical standard curve). Determine the equation and  $R^2$  value of the resulting trend line.
3. Suppose the equation of the trend line of the standard curve is  $y = Ax + B$ , calculate the choline concentration of test samples as follows:

$$[\text{Choline}] = \frac{\Delta\text{OD}_{550\text{nm}} - B}{A}$$

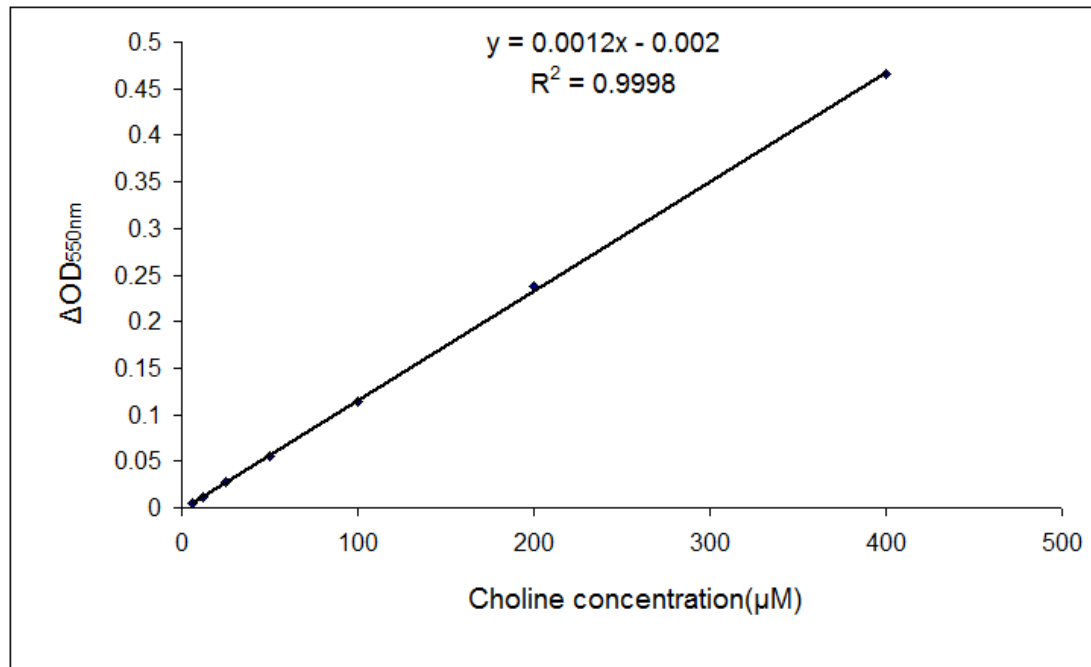


Figure1. A typical choline standard curve measured by ScienCell™ Choline Assay kit.