

**Human TGF α ELISA Kit
(hTGF- α -ELISA)***Cat. No. EK0511**96 Tests in 8 x 12 divisible strips***Background**

Transforming growth factor alpha (TGF- α) is upregulated in some human cancers. It is produced in macrophages, brain cells, and keratinocytes, and it induces epithelial development. It is closely related to epidermal growth factor (EGF) and can also bind to the EGF receptor (EGFR) with similar effects. TGF- α stimulates neural cell proliferation in the adult injured brain. Transforming growth factor alpha gene (TGFA) maps to human chromosome 2 close to the breakpoint of the t(2;8) variant translocation in Burkitt lymphoma. Synthetic TGF- α was as active as murine EGF in binding to the EGFR and in stimulation of anchorage-dependent and -independent growth of normal indicator cells in culture. Synthetic TGF- α stimulates plasminogen activator production in A 431 and HeLa cells; the stimulation was similar to that induced by EGF. Furthermore, synthetic human TGF- α showed similar immunoreactivity when compared with rat TGF- α . Thus, the 50-amino acid TGF- α is likely to be the bioactive principle produced and secreted by tumor cell lines.

ScienCell's human TGF- α ELISA Kit is based on standard sandwich enzyme-linked immune-sorbent assay technology. Human TGF- α -specific polyclonal antibodies are pre-coated onto 8 x 12 divisible strips. The human specific detection polyclonal antibodies are biotinylated. The test samples and biotinylated detection antibodies are subsequently added to the wells and then washed with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex is added and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB is used to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product that changes into yellow after adding acidic stop solution. The intensity of yellow is proportional to the amount of human TGF- α captured in strips.

Size	96 Tests in 8 x 12 divisible strips
Assay type	Sandwich ELISA
Range	15.6 pg/ml- 1000 pg/ml
Sensitivity	< 1 pg/ml
Specificity	Cross-reactivates with TGF β 2, TGF β 3, TGF β 5 <1%.
Storage	Store at 4°C for frequent use, at -20°C for infrequent use. Avoid multiple freeze-thaw cycles
Shipping	Shipped on gel ice.

Expiration	Four months at 4°C and eight months at -20°C.
Application	For quantitative detection of human TGF α in serum, plasma, body fluids, tissue lysates or cell culture supernatants.
Kit components	<ol style="list-style-type: none"> 1. Lyophilized recombinant human TGF-α standard: 10 ng/tube\times2. 2. 8 x 12 divisible strips pre-coated with anti- human TGFα antibody. 3. Sample diluent buffer: 30 ml 4. Biotinylated anti- human TGFα antibody: 130μl, dilution 1:100. 5. Antibody diluent buffer: 12ml. 6. Avidin-Biotin-Peroxidase Complex (ABC): 130μl, dilution 1:100. 7. ABC diluent buffer: 12ml. 8. TMB color developing agent: 10ml. 9. TMB stop solution: 10ml.
Materials	1. Microplate reader.
Required But	2. Automated plate washer.
Not Provided	<ol style="list-style-type: none"> 3. Adjustable pipettes and pipette tips. Multi-channel pipettes are recommended for large number of samples. 4. Clean tubes and Eppendorf tubes. 5. Washing buffer (neutral PBS or TBS). Preparation of 0.01M TBS: Add 1.2g Tris, 8.5g Nacl; 450μl of purified acetic acid or 700μl of concentrated hydrochloric acid to 1000ml H₂O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L. Preparation of 0.01 M PBS: Add 8.5g sodium chloride, 1.4g Na₂HPO₄ and 0.2g NaH₂PO₄ to 1000ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.
Usage	This product is for research use only. It is not approved for use in humans, animals, or <i>in vitro</i> diagnostic procedures.

Reference

1. In vivo induction of massive proliferation, directed migration, and differentiation of neural cells in the adult mammalian brain. Proc Natl Acad Sci USA. 2000 Dec 19; 97(26): 14686-91;
2. Brissenden, J. E., Derynck, R., Francke, U. Transforming growth factor alpha gene (TGFA) maps to human chromosome 2 close to the breakpoint of the t(2;8) variant translocation in Burkitt lymphoma. Cytogenet. Cell Genet. 40: 589-only, 1985.
3. Tam, J. P., Scheikh, M. A., Solomon, D. S., Ossowski, L. Efficient synthesis of human type alpha transforming growth factor: its physical and biological characterization. Proc. Nat. Acad. Sci. 83: 8082-8086, 1986.

Protocol for Human TGF α ELISA (96 well format)

Notes before you begin

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, a pilot experiment using standards and a small number of samples is recommended.
2. The TMB Color developing agent should be colorless and transparent before using.
3. Before using the kit, spin tubes and bring down all components to the bottom of tubes.

4. A duplicate well assay is recommended for both standard and samples.
5. Do not let strips dry, as this will inactivate active components in wells.
6. Do not reuse tips and tubes to avoid cross contamination.
7. Avoid using reagents from different batches.
8. In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37°C for 30 minutes before use.

Preparation

Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C.

Avoid repeated freeze-thaw cycles.

- **Cell culture supernatants, tissue lysate or body fluids:** Remove particulates by centrifugation, assay immediately or aliquot and store samples at -20°C.
- **Serum:** Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 10 minutes. Analyze the serum immediately or aliquot and store frozen at -70°C.
- **Plasma:** Collect plasma using EDTA as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. For eliminating platelet, suggesting that further centrifugation for 10 minutes at 2-8°C at 10000 x g. Analyze immediately or aliquot and store frozen at -70°C. Heparin and citrate are not recommended as the anticoagulant.

Note: Bovine serum used in cell culture supernatants may contain TGF α , avoiding using it.

Sample Dilution Guideline

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. **The sample must be well mixed with the diluents buffer.**

- **High target protein concentration (10-100 ng/ml).** The working dilution is 1:100. i.e. Add 1 μ l sample into 99 μ l sample diluent buffer.
- **Medium target protein concentration (1-10 ng/ml).** The working dilution is 1:10. i.e. Add 10 μ l sample into 90 μ l sample diluent buffer.
- **Low target protein concentration (15.6-1000 pg/ml).** The working dilution is 1:2. i.e. Add 50 μ l sample to 50 μ l sample diluent buffer.
- **Very Low target protein concentration (\leq 15.6 pg/ml).** No dilution necessary, or the working dilution is 1:2.

Reagent Preparation and Storage

A. Reconstitution of the human TGF α standard: TGF α standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of TGF α standard (10 ng per tube) are included in each kit. Use one tube for each experiment.

- 10,000 pg/ml of human TGF α standard solution: Add 1 ml sample diluent buffer into one tube, keep the tube at room temperature for 10 minutes and mix thoroughly.
- 1000 pg/ml of human TGF α standard solution: Add 0.1 ml of the above 10ng/ml TGF α standard solution into 0.9 ml sample diluent buffer and mix thoroughly.
- 500 pg/ml \rightarrow 15.6 pg/ml of human TGF α standard solutions: Label 6 Eppendorf tubes with 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml, 31.2pg/ml, 15.6pg/ml respectively. Aliquot 0.3 ml of the sample diluent buffer into each tube. Add 0.3 ml of the above 1000pg/ml TGF α standard solution into 1st tube and mix.

Transfer 0.3 ml from 1st tube to 2nd tube and mix. Transfer 0.3 ml from 2nd tube to 3rd tube and mix, and so on.

Note: The standard solutions are best used within 2 hours. The 10 ng/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

B. Preparation of biotinylated anti-human TGF α antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume)
- Biotinylated anti-human TGF α antibody should be diluted in 1:100 with the antibody diluent buffer and mixed thoroughly.

C. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume)
- Avidin- Biotin-Peroxidase Complex (ABC) should be diluted in 1:100 with the ABC dilution buffer and mixed thoroughly.

Assay Procedure

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 minutes before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard TGF α detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of TGF α amount in samples.

1. Aliquot 0.1ml per well of the 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml, 31.2pg/ml, 15.6pg/ml human TGF α standard solutions into the pre-coated strips. Add 0.1ml of the sample diluent buffer into the control well (**blank well**). Add 0.1ml of each properly diluted sample of human serum, plasma, body fluids, tissue lysates or cell culture supernatants to each empty well. See “**Sample Dilution Guideline**” above for details. We recommend that each human TGF α standard solution and each sample is measured in duplicate.
2. Seal the strips with the cover and incubate at 37°C for 90 minutes.
3. Remove the cover, discard strips’ contents, and blot the strips onto paper towels or other absorbent material. **Do NOT** let the wells completely dry at any time.
4. Add 0.1ml of biotinylated anti-human TGF α antibody working solution into each well and incubate the strips at 37°C for 60 minutes.
5. Wash strips 3 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1 minute. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (**Strips Washing Method:** Discard the solution in the strips without touching the side walls. Blot the strips onto paper towels or other absorbent material. Soak each well with at least 0.3 ml PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of **THREE** washes. Note: For automated washing, aspirate all wells and wash **THREE** times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the strips onto paper towels or other absorbent material).
6. Add 0.1ml of prepared ABC working solution into each well and incubate the strips at 37°C for 30 minutes.
7. Wash strips 5 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1-2 minutes. Discard the washing buffer and blot the strips onto paper towels or other absorbent material.(See Step 5 for strips washing method).
8. Add 90 μ l of prepared TMB color developing agent into each well and incubate strips at 37°C in dark for 15-20 minutes (**Note:** For reference only, the optimal incubation time should be determined by end user. And the shades

of blue can be seen in the wells with the four most concentrated human TGF α standard solutions; the other wells show no obvious color).

9. Add 0.1ml of prepared TMB stop solution into each well. The color changes into yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 minutes after adding the stop solution.

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of blank well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The human TGF α concentration of the samples can be interpolated from the standard curve.

Note: if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

Summary

1. Add samples and standards and incubate the strips at 37°C for 90 minutes. Do not wash.
2. Add biotinylated antibodies and incubate the strips at 37°C for 60 minutes. Wash strips 3 times with 0.01M TBS.
3. Add ABC working solution and incubate the strips at 37°C for 30 minutes. Wash strips 5 times with 0.01M TBS.
4. Add TMB color developing agent and incubate the strips at 37°C in dark for 15-20 minutes.
5. Add TMB stop solution and read.

Typical Data Obtained from Human TGF α (TMB reaction incubate at 37°C for 15 minutes)

Concentration (pg/ml)	0.0	15.6	31.3	62.5	125	250	500	1000
Absorbance (450 nm)	0.064	0.125	0.177	0.365	0.905	1.329	2.322	2.902

Typical Human TGF α ELISA Kit Standard Curve

This standard curve was generated for demonstration purpose only. A standard curve must be run with each assay.

