

Mouse TNF α ELISA Kit
(mTNF-ELISA)

Cat. No. EK0527

96 Tests in 8 x 12 divisible strips

Background

Tumor necrosis factor α (TNF α or TNF) is secreted by macrophages in response to inflammation, infection, and cancer. Mouse Tumor Necrosis Factor (TNF) and Lymphotoxin (TNF β) are cytotoxic proteins which have similar biological activities and share 30% amino acid homology. TNF α is produced by monocytes, which can stimulate endothelial cells to produce the multi-lineage growth factor granulocyte-macrophage colony-stimulating factor (GM-CSF) and extend the role of this immunoregulatory protein to the regulation of hematopoiesis *in vitro*. TNF is a soluble protein that causes damage to tumor cells, but has no effect on normal cells. Mouse TNF has been purified to apparent homogeneity as a 17.3 kDa protein from HL-60 leukemia cells and has shown cytotoxic and cytostatic activities against various mouse tumor cell lines. The mouse TNF cDNA is 1585 base pairs in length and encodes a protein of 233 amino acids. The mature protein begins at residue 77, leaving a long leader sequence of 76 amino acids. TNF α has been mapped to human chromosome 6.

ScienCell's mouse TNF α ELISA Kit is based on standard sandwich enzyme-linked immune-sorbent assay technology. Mouse TNF α -specific monoclonal antibodies are pre-coated onto 8 x 12 strips. The mouse-specific detection polyclonal antibodies are biotinylated. The test samples and biotinylated detection antibodies are subsequently added to the wells and then washed with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex is added and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB is used to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product that changes to yellow after adding acidic stop solution. The intensity of yellow is proportional to the amount of mouse TNF α in the sample that is captured on the strips.

Size

96 Tests in 8 x 12 divisible strips

Assay type

Sandwich ELISA

Range

15.6 pg/ml-1000 pg/ml (Body fluids, tissue lysates or cell culture supernatants)
7.8 pg/ml – 500 pg/ml (Mouse serum)

Sensitivity

< 1 pg/ml

Specificity

No detectable cross-reactivity with any other cytokine.

Storage	Store at 4°C for frequent use, at -20°C for infrequent use. Avoid multiple freeze-thaw cycles.
Shipping	Shipped on gel ice.
Expiration	Four months at 4°C and eight months at -20°C.
Application	For quantitative detection of mouse TNF α in serum, plasma, body fluids, tissue lysates or cell culture supernatants.
Kit components	<ol style="list-style-type: none"> 1. Lyophilized recombinant mouse TNFα standard: 10 ng/tube\times2. 2. 8 x 12 divisible strips pre-coated with anti- mouse TNFα antibody. 3. Sample diluent buffer: 30 ml 4. Biotinylated anti-mouse TNFα antibody: 130μl, dilution 1:100. 5. Antibody diluent buffer: 12ml. 6. Avidin-Biotin-Peroxidase Complex (ABC): 130μl, dilution 1:100. 7. ABC diluent buffer: 12 ml 8. TMB color developing agent: 10 ml. 9. TMB stop solution: 10 ml.
Materials Required But Not Provided	<ol style="list-style-type: none"> 1. Microplate reader. 2. Automated plate washer. 3. Adjustable pipettes and pipette tips. Multichannel pipettes are recommended for large number of samples. 4. Clean tubes and Eppendorf tubes. 5. Washing buffer (neutral PBS or TBS). <p>Preparation of 0.01M TBS: Add 1.2g Tris, 8.5g NaCl; 450μl of purified acetic acid or 700μl of concentrated hydrochloric acid to 1000ml H₂O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.</p> <p>Preparation of 0.01 M PBS: Add 8.5g NaCl, 1.4g Na₂HPO₄ and 0.2g NaH₂PO₄ to 1000ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.</p>
Usage	This product is for research use only. It is not approved for use in humans, animals, or <i>in vitro</i> diagnostic procedures.

Reference

1. Brenner, D. A.; O'Hara, M.; Angel, P.; Chojkier, M.; Karin, M. Prolonged activation of JUN and collagenase genes by tumour necrosis factor α . *Nature* 337: 661-663, 1989.
2. Nedwin, G. E.; Naylor, S. L.; Sakaguchi, A. Y.; Smith, D.; Jarrett-Nedwin, J.; Pennica, D.; Goeddel, D. V.; Gray, P. W. Human lymphotoxin and tumor necrosis factor genes: structure, homology and chromosomal localization. *Nucleic Acids Res.* 13: 6361-6373, 1985.
3. Broudy, V. C.; Kaushansky, K.; Segal, G. M.; Harlan, J. M.; Adamson, J. W. Tumor necrosis factor type α stimulates human endothelial cells to produce granulocyte/macrophage colony-stimulating factor. *Proc. Nat. Acad. Sci.* 83: 7467-7471, 1986.
4. Wang, A. M.; Creasey, A. A.; Ladner, M. B.; Lin, L. S.; Strickler, J.; Van Arsdel, J. N.; Yamamoto, R.; Mark, D. F. Molecular cloning of the complementary DNA for human tumor necrosis factor. *Science* 228: 149-154, 1985.

Protocol for Mouse TNF α ELISA (96-well format)

Notes before you begin

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, a pilot experiment using standards and a small number of samples is recommended.
2. The TMB Color developing agent should be colorless and transparent before using.
3. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
4. A duplicate well assay is recommended for both standard and samples.
5. Do not let wells dry, as this will inactivate active components in wells.
6. Do not reuse tips and tubes to avoid cross contamination.
7. Avoid using reagents from different batches.
8. In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37°C for 30 minutes before use.

Preparation

Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C.

Avoid repeated freeze-thaw cycles.

- **Cell culture supernatants, tissue lysate, or body fluids:** Remove particulates by centrifugation, assay immediately or aliquot and store samples at -20°C.
- **Serum:** Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature or coat at 4°C overnight. Centrifuge at approximately 2000 X g for 20 minutes. Analyze the serum immediately or aliquot and store frozen at -20°C.
- **Plasma:** Collect plasma using heparin, EDTA, citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Analyze immediately or aliquot and store frozen at -20°C.

Note: The sample with hyperlipidemia and haemolyticus is not suitable for this kit.

Sample Dilution Guideline

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. **The sample must be mixed well with the diluent buffer.**

- **High target protein concentration (10-100 ng/ml).** The working dilution is 1:100. i.e. Add 1 μ l sample into 99 μ l sample diluent buffer.
- **Medium target protein concentration (1-10 ng/ml).** The working dilution is 1:10. i.e. Add 10 μ l sample into 90 μ l sample diluent buffer.
- **Low target protein concentration (15.6-1000 pg/ml).** The working dilution is 1:2. i.e. Add 50 μ l sample to 50 μ l sample diluent buffer.
- **Very Low target protein concentration (\leq 15.6 pg/ml).** No dilution necessary, or the working dilution is 1:2.

Reagent Preparation and Storage

- A. Reconstitution of the mouse TNF α standard: TNF α standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of TNF α standard (10 ng per tube) are included in each kit. Use one tube for each experiment.

- 10,000 pg/ml of mouse TNF α standard solution: Add 1 ml sample diluent buffer into one tube, keep the tube at room temperature for 10 minutes and mix thoroughly.
- 1000 pg/ml of mouse TNF α standard solution: Add 0.1 ml of the above 10ng/ml TNF α standard solution into 0.9 ml sample diluent buffer and mix thoroughly.
- 500 pg/ml→15.6 pg/ml of mouse TNF α standard solutions: Label 6 Eppendorf tubes with 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml, 31.3pg/ml, 15.6pg/ml, respectively. Aliquot 0.3 ml of the sample diluent buffer into each tube. Add 0.3 ml of the above 1000pg/ml TNF α standard solution into 1st tube and mix. Transfer 0.3 ml from 1st tube to 2nd tube and mix. Transfer 0.3 ml from 2nd tube to 3rd tube and mix, and so on.

Note: The standard solutions are best used within 2 hours. The 10 ng/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

B. Preparation of biotinylated anti-mouse TNF α antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).
- Biotinylated anti-mouse TNF α antibody should be diluted in 1:100 with the antibody diluent buffer and mixed thoroughly.

C. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).
- Avidin-Biotin-Peroxidase Complex (ABC) should be diluted in 1:100 with the ABC dilution buffer and mixed thoroughly.

Assay Procedure

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 minutes before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard TNF α detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of TNF α amount in samples.

1. Aliquot 0.1ml per well of the 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml, 31.3pg/ml, 15.6pg/ml mouse TNF α standard solutions into the pre-coated 8 x 12 divisible strips. Add 0.1ml of the sample diluent buffer into the control well (**blank well**). Add 0.1ml of each properly diluted sample of mouse serum, plasma, body fluids, tissue lysates or cell culture supernatants to each empty well. See “**Sample Dilution Guideline**” above for details. We recommend that each mouse TNF α standard solution and each sample is measured in duplicate.
2. Seal the strips with the cover and incubate at 37°C for 90 minutes.
3. Remove the cover, discard strips’ contents, and blot the strips onto paper towels or other absorbent material. **Do NOT** let the wells completely dry at any time.
4. Add 0.1ml of biotinylated anti-mouse TNF α antibody working solution into each well and incubate the strips at 37°C for 60 minutes.
5. Wash the strips 3 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1 minute. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (**Strips Washing Method:** Discard the solution in the wells without touching the side walls. Blot the strips onto paper towels or other absorbent material. Soak each well with at least 0.3 ml PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of **THREE** washes. Note: For automated washing, aspirate all wells and wash **THREE** times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the strips onto paper towels or other absorbent material).

6. Add 0.1ml of prepared ABC working solution into each well and incubate the strips at 37°C for 30 minutes.
7. Wash the strips 5 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1-2 minutes. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (See Step 5 for strips washing method).
8. Add 90 µl of prepared TMB color developing agent into each well and incubate the strips at 37°C in dark for 20-25 minutes (**Note:** For reference only, the optimal incubation time should be determined by end user. And the shades of blue can be seen in the wells with the four most concentrated mouse TNFα standard solutions; the other wells show no obvious color).
9. Add 0.1ml of prepared TMB stop solution into each well. The color changes to yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 minutes after adding the stop solution.

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of blank well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The mouse TNFα concentration of the samples can be interpolated from the standard curve.

Note: if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

Summary

1. Add samples and standards and incubate the strips at 37°C for 90 minutes. Do not wash.
2. Add biotinylated antibodies and incubate the strips at 37°C for 60 minutes. Wash strips 3 times with 0.01M TBS.
3. Add ABC working solution and incubate the strips at 37°C for 30 minutes. Wash strips 5 times with 0.01M TBS.
4. Add TMB color developing agent and incubate the strips at 37°C in dark for 20-25 minutes.
5. Add TMB stop solution and read.

Typical Data Obtained from Mouse TNFα

(TMB reaction incubate at 37°C for 22 minutes)

Concentration (pg/ml)	0.0	15.6	31.3	62.5	125	250	500	1000
Absorbance (450 nm)	0.035	0.076	0.113	0.180	0.339	0.861	1.344	2.394

Typical Mouse TNFα ELISA Kit Standard Curve

This standard curve was generated for demonstration purpose only. A standard curve must be run with each assay.

