

Mouse P-Cadherin ELISA Kit
(mPCad-ELISA)*Cat. No. EK0668**96 Tests in 8 x 12 divisible strips*

Background P-Cadherin, also known as Cadherin-3 (CDH3), is a protein that in humans is encoded by the CDH3 gene. This gene is a classical cadherin from the cadherin superfamily. This gene is located in a six-cadherin cluster in a region on the long arm of chromosome 16. It is involved in the loss of heterozygosity events in breast and prostate cancer. In addition, aberrant expression of this protein is observed in cervical adenocarcinomas. Mutations in this gene have been associated with congenital hypotrichosis and juvenile macular dystrophy. Cells expressing CDH3 also adhere to one another more tightly than the parental cell line.

ScienCell's mouse P-Cadherin ELISA kit is based on standard sandwich enzyme-linked immune-sorbent assay technology. Mouse P-Cadherin-specific polyclonal antibodies are pre-coated onto 8 x 12 divisible strips. The mouse-specific detection monoclonal antibodies are biotinylated. The test samples and biotinylated detection antibodies are subsequently added to the wells and then washed with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex is added and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB is used to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product that changes to yellow after adding acidic stop solution. The intensity of yellow is proportional to the amount of mouse P-Cadherin in the sample that is captured on the strips.

Size	96 Tests in 8 x 12 divisible strips
Assay type	Sandwich ELISA
Range	62.5 pg/ml- 4000 pg/ml
Sensitivity	< 10 pg/ml
Specificity	Natural and recombinant mouse P-Cadherin
Cross-reactivity	No detectable cross-reactivity with other relevant proteins
Storage	Store at 4°C for 6 months, at -20°C for 12 months. Avoid multiple freeze-thaw cycles.
Shipping	Shipped on gel ice.
Application	For quantitative detection of mouse P-Cadherin in cell culture supernatants and serum.

Kit components	1. Lyophilized recombinant mouse P-Cadherin standard: 10 ng/tube×2. 2. 8 x 12 divisible strips pre-coated with anti- mouse P-Cadherin antibody. 3. Sample diluent buffer: 30 ml 4. Biotinylated anti-mouse P-Cadherin antibody: 130µl, dilution 1:100. 5. Antibody diluent buffer: 12ml. 6. Avidin-Biotin-Peroxidase Complex (ABC): 130µl, dilution 1:100. 7. ABC diluent buffer: 12 ml 8. TMB color developing agent: 10 ml. 9. TMB stop solution: 10 ml.
Materials	1. Microplate reader.
Required But	2. Automated plate washer.
Not Provided	3. Adjustable pipettes and pipette tips. Multichannel pipettes are recommended for large amount of samples. 4. Clean tubes and Eppendorf tubes. 5. Washing buffer (neutral PBS or TBS). Preparation of 0.01M TBS: Add 1.2g Tris, 8.5g NaCl; 450µl of purified acetic acid or 700µl of concentrated hydrochloric acid to 1000ml H₂O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L. Preparation of 0.01 M PBS: Add 8.5g NaCl, 1.4g Na₂HPO₄ and 0.2g NaH₂PO₄ to 1000ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.
Usage	This product is for research use only. It is not approved for use in humans, animals, or <i>in vitro</i> diagnostic procedures.

Reference

1. Kaupmann K, Becker-Follmann J, Scherer G, Jockusch H, Starzinski-Powitz A (December 1992). "The gene for the cell adhesion molecule M-cadherin maps to mouse chromosome 8 and human chromosome 16q24.1-qter and is near the E-cadherin (uvomorulin) locus in both species". Genomics 14 (2): 488–90.
2. "Entrez Gene: CDH3 cadherin 3, type 1, P-cadherin (placental)"
3. Shimoyama, Y., Yoshida, T., Terada, M., Shimosato, Y., Abe, O., Hirohashi, S. Molecular cloning of a human Ca(2+)-dependent cell-cell adhesion molecule homologous to mouse placental cadherin: its low expression in human placental tissues. J. Cell Biol. 109: 1787-1794, 1989.

Protocol for Mouse P-Cadherin ELISA (96-well format)

Notes before you begin

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, a pilot experiment using standards and a small number of samples is recommended.
2. The TMB Color developing agent should be colorless and transparent before using.
3. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
4. A duplicate well assay is recommended for both standard and samples.
5. Do not let wells dry, as this will inactivate active components in wells.
6. Do not reuse tips and tubes to avoid cross contamination.
7. Avoid using reagents from different batches.

8. In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37°C for 30 minutes before use.

Preparation

Sample Preparation and Storage

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C.

Avoid repeated freeze-thaw cycles.

- **Cell culture supernatants:** Remove particulates by centrifugation, assay immediately or aliquot and store samples at -20°C.
- **Serum:** Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 minutes. Analyze the serum immediately or aliquot and store frozen at -20°C.

Sample Dilution Guideline

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. **The sample must be mixed well with the diluent buffer.**

- **High target protein concentration (40-400 ng/ml).** The working dilution is 1:100. i.e. Add 1 µl sample into 99 µl sample diluent buffer.
- **Medium target protein concentration (4-40 ng/ml).** The working dilution is 1:10. i.e. Add 10 µl sample into 90 µl sample diluent buffer.
- **Low target protein concentration (62.5-4000 pg/ml).** The working dilution is 1:2. i.e. Add 50 µl sample to 50 µl sample diluent buffer.
- **Very Low target protein concentration (≤ 62.5 pg/ml).** No dilution necessary, or the working dilution is 1:2.

Reagent Preparation and Storage

A. Reconstitution of the mouse P-Cadherin standard: P-Cadherin standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of P-Cadherin standard (10 ng per tube) are included in each kit. Use one tube for each experiment.

- 10,000 pg/ml of mouse P-Cadherin standard solution: Add 1 ml sample diluent buffer into one tube, keep the tube at room temperature for 10 minutes and mix thoroughly.
- 4000 pg/ml of mouse P-Cadherin standard solution: Add 0.4ml of the above 10ng/ml P-Cadherin standard solution into 0.6ml sample diluent buffer and mix thoroughly.
- 2000 pg/ml→62.5 pg/ml of mouse P-Cadherin standard solutions: Label 6 Eppendorf tubes with 2000 pg/ml, 1000 pg/ml, 500 pg/ml, 250 pg/ml, 125 pg/ml, 62.5 pg/ml respectively. Aliquot 0.3ml of the sample diluent buffer into each tube. Add 0.3ml of the above 4000 pg/ml P-Cadherin standard solution into 1st tube and mix. Transfer 0.3 ml from 1st tube to 2nd tube and mix. Transfer 0.3ml from 2nd tube to 3rd tube and mix, and so on.

Note: The standard solutions are best used within 2 hours. The 10 ng/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

B. Preparation of biotinylated anti-mouse P-Cadherin antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).

- Biotinylated anti-mouse P-Cadherin antibody should be diluted in 1:100 with the antibody diluent buffer and mixed thoroughly.
- C. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.
- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).
 - Avidin-Biotin-Peroxidase Complex (ABC) should be diluted in 1:100 with the ABC dilution buffer and mixed thoroughly.

Assay Procedure

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 minutes before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard P-Cadherin detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of P-Cadherin amount in samples.

1. Aliquot 0.1ml per well of the 4000pg/ml, 2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml mouse P-Cadherin standard solutions into the pre-coated 8 x 12 divisible strips. Add 0.1ml of the sample diluent buffer into the control well (**blank well**). Add 0.1ml of each properly diluted sample of mouse cell culture supernatants or serum to each empty well. See “**Sample Dilution Guideline**” above for details. We recommend that each mouse P-Cadherin standard solution and each sample is measured in duplicate.
2. Seal the strips with the cover and incubate at 37°C for 90 minutes.
3. Remove the cover, discard the strips’ contents, and blot the strips onto paper towels or other absorbent material. **Do NOT** let the wells completely dry at any time.
4. Add 0.1ml of biotinylated anti-mouse P-Cadherin antibody working solution into each well and incubate the strips at 37°C for 60 minutes.
5. Wash the strips 3 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1 minute. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (**Strips Washing Method:** Discard the solution in the wells without touching the side walls. Blot the strips onto paper towels or other absorbent material. Soak each well with at least 0.3 ml PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of THREE washes. Note: For automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the strips onto paper towels or other absorbent material).
6. Add 0.1ml of prepared ABC working solution into each well and incubate the strips at 37°C for 30 minutes.
7. Wash the strips 5 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1-2 minutes. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (See Step 5 for strip washing method).
8. Add 90 µl of prepared TMB color developing agent into each well and incubate the strips at 37°C in dark for 20-25 minutes (**Note:** For reference only, the optimal incubation time should be determined by end user. And the shades of blue can be seen in the wells with the four most concentrated mouse P-Cadherin standard solutions; the other wells show no obvious color).
9. Add 0.1ml of prepared TMB stop solution into each well. The color changes to yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 minutes after adding the stop solution.

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of blank well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The mouse P-Cadherin concentration of the samples can be interpolated from the standard curve.

Note: if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

Summary

1. Add samples and standards and incubate the strips at 37°C for 90 minutes. Do not wash.
2. Add biotinylated antibodies and incubate the strips at 37°C for 60 minutes. Wash strips 3 times with 0.01M TBS.
3. Add ABC working solution and incubate the strips at 37°C for 30 minutes. Wash strips 5 times with 0.01M TBS.
4. Add TMB color developing agent and incubate the strips at 37°C in dark for 20-25 minutes.
5. Add TMB stop solution and read.

Typical Data Obtained from Mouse P-Cadherin

(TMB reaction incubate at 37°C for 20 minutes)

Concentration (pg/ml)	0.0	62.5	125	250	500	1000	2000	4000
Absorbance (450 nm)	0.036	0.082	0.111	0.173	0.303	0.613	1.095	2.050

Typical Mouse P-Cadherin ELISA Kit Standard Curve

This standard curve was generated for demonstration purpose only. A standard curve must be run with each assay.

