



## Mouse Perineurial Fibroblasts (MPNF)

Catalog #M1710-57

### Cell Specification

Perineurial fibroblasts are of mesenchymal origin and form the perineurium. The perineurium plays an important role in maintaining the integrity of the internal peripheral nerve environment by creating a physical barrier that, under physiologic conditions, limits the entry of biologically active proteins, infectious agents, and blood-borne cells into the nerve bundles [1]. The perineurial fibroblasts are characterized by distinct ultrastructural features, including non-branching thin cytoplasmic processes coated by an external lamina and joined at their ends by a tight junction, few organelles, actin and vimentin filaments, and numerous pinocytotic vesicles [2]. Perineurial fibroblasts are initially recruited from the surrounding mesenchyme to form a loose, permeable sheath around axons and Schwann cells, where they are separated by the extracellular matrix. These cells later undergo a mesenchymal-to-epithelial transition to form tight junctions and organize into the perineurium. Perineurial fibroblasts are immunoreactive for vimentin and epithelial membrane antigen but not for the Schwann cell marker S-100 [3].

MPNF from ScienCell Research Laboratories are isolated from postnatal day 8 C57BL/6 mouse sciatic nerve. MPNF are cryopreserved at passage one and delivered frozen. Each vial contains  $>5 \times 10^5$  cells in 1 ml volume. MPNF are characterized by immunofluorescence with antibodies specific to vimentin and fibronectin. MPNF are negative for mycoplasma, bacteria, yeast, and fungi. MPNF are guaranteed to further expand for 5 population doublings under the conditions provided by ScienCell Research Laboratories.

### Recommended Medium

It is recommended to use Fibroblast Medium (FM, Cat. #2301) for culturing MPNF *in vitro*.

### Product Use

MPNF are for research use only. They are not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

### Storage

Upon receiving, directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

### Shipping

Dry ice.

### References

- [1] Salzer JL. (1999) "Creating barriers: a new role for Schwann cells and desert hedgehog." *Neuron*. 22: 627-9.
- [2] Erlandson RA. (1991) "The enigmatic perineurial cell and its participation in tumors and in tumor like entities." *Ultrastruct Pathol*. 15: 335-51.
- [3] Ariza A, Bilbao JM, Rosai J. (1988) "Immunohistochemical detection of epithelial membrane antigen in normal and perineurial cells and perineurioma." *Am J Surg Pathol*. 12: 678-83.

## Instructions for culturing cells

---

**Caution:** Cryopreserved cells are very delicate. Thaw the vial in a 37°C water bath and return the cells to culture as quickly as possible with minimal handling!

**Note:** ScienCell primary cells must be cultured in a 37°C, 5% CO<sub>2</sub> incubator. Cells are only warranted if ScienCell media and reagents are used and the recommended protocols are followed.

### Initiating the culture:

1. Prepare a poly-L-lysine-coated culture vessel (2 µg/cm<sup>2</sup>, T-75 flask is recommended). Add 10 ml of sterile water to a T-75 flask and then add 15 µl of poly-L-lysine stock solution (10 mg/ml, Cat. #0413). Leave the vessel in a 37°C incubator overnight (or for a minimum of one hour).
2. Prepare complete medium. Decontaminate the external surfaces of medium bottle and medium supplement tubes with 70% ethanol and transfer them to a sterile field. Aseptically transfer supplement to the basal medium with a pipette. Rinse the supplement tube with medium to recover the entire volume.
3. Rinse the poly-L-lysine-coated vessel twice with sterile water and then add 15 ml of complete medium. Leave the vessel in the sterile field and proceed to thaw the cryopreserved cells.
4. Place the frozen vial in a 37°C water bath. Hold and rotate the vial gently until the contents completely thaw. Promptly remove the vial from the water bath, wipe it down with 70% ethanol, and transfer it to the sterile field.
5. Carefully remove the cap without touching the interior threads. Gently resuspend and dispense the contents of the vial into the equilibrated, poly-L-lysine-coated culture vessel. A seeding density of 5,000 cells/cm<sup>2</sup> is recommended.

*Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to the cells than the effect of residual DMSO in the culture. It is also important that cells are plated in poly-L-lysine-coated culture vessels to promote cell attachment.*

6. Replace the cap or lid of the culture vessel and gently rock the vessel to distribute the cells evenly. Loosen cap, if necessary, to allow gas exchange.
7. Return the culture vessel to the incubator.
8. For best results, do not disturb the culture for at least 16 hours after the culture has been initiated. Refresh culture medium the next day to remove residual DMSO and unattached cells, then every other day thereafter.

### Maintaining the culture:

1. Refresh supplemented culture medium the next morning after establishing a culture from cryopreserved cells.
2. Change the medium every three days thereafter, until the culture is approximately 70% confluent.

3. Once the culture reaches 70% confluency, change medium every other day until the culture is approximately 90% confluent.

### **Subculturing:**

1. Subculture when the culture reaches 90% confluency or above.
2. Prepare poly-L-lysine-coated culture vessels ( $2 \mu\text{g}/\text{cm}^2$ ) one day before subculture.
3. Warm complete medium, trypsin/EDTA solution (T/E, Cat. #0103), T/E neutralization solution (TNS, Cat. #0113), and DPBS ( $\text{Ca}^{++}$ - and  $\text{Mg}^{++}$ -free, Cat. #0303) to **room temperature**. We do not recommend warming reagents and medium in a  $37^\circ\text{C}$  water bath prior to use.
4. Rinse the cells with DPBS.
5. Add 8 ml of DPBS and then 2 ml of T/E solution into flask (in the case of a T-75 flask). Gently rock the flask to ensure complete coverage of cells by T/E solution. Incubate the flask in a  $37^\circ\text{C}$  incubator for 1 to 2 minutes or until cells completely round up. Use a microscope to monitor the change in cell morphology.
6. During incubation, prepare a 50 ml conical centrifuge tube with 5 ml of fetal bovine serum (FBS, Cat. #0500).
7. Transfer T/E solution from the flask to the 50 ml centrifuge tube (a small percent of cells may detach) and continue to incubate the flask at  $37^\circ\text{C}$  for another 1 to 2 minutes (no solution in the flask at this moment).
8. At the end of incubation, gently tap the side of the flask to dislodge cells from the surface. Check under a microscope to make sure that all cells detach.
9. Add 5 ml of TNS solution to the flask and transfer detached cells to the 50 ml centrifuge tube. Rinse the flask with another 5 ml of TNS to collect the residual cells.
10. Examine the flask under a microscope for a successful cell harvest by looking at the number of cells being left behind; there should be less than 5%.

*Note: Use ScienCell T/E solution that is optimized to minimize cell damages due to over trypsinization.*

11. Centrifuge the 50 ml centrifuge tube at 1000 rpm for 5 minutes. Resuspend cells in culture medium.
12. Count and plate cells in a new poly-L-lysine-coated culture vessel with the recommended cell density.

**Note:** We do not recommend cryopreservation of primary cells by the end user. Refreezing cells may damage them and affect cell performance. ScienCell does not guarantee primary cells cryopreserved by the end user.

*Caution: Handling animal-derived products is potentially biohazardous. Always wear gloves and safety glasses when working with these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].*

[1] Grizzle WE, Polt S. (1988) "Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues." *J Tissue Cult Methods*. 11: 191-9.